## academicJournals

Vol. 8(12), pp. 537-545, December 2014 DOI: 10.5897/AJPS2014.1221 Article Number: 5D100A149355 ISSN 1996-0824 Copyright © 2014 Author(s) retain the copyright of this article http://www.academicjournals.org/AJPS

African Journal of Plant Science

Review

# Distribution, chemical composition and medicinal importance of saffron (*Crocus sativus* L.)

Sabbi Jan<sup>1</sup>\*, Aijaz A. Wani<sup>1</sup>, Azra N. Kamili<sup>2</sup> and Mahpara Kashtwari<sup>1</sup>

<sup>1</sup>Cytogenetics and Reproductive Biology Laboratory, Department of Botany, University of Kashmir, 190 006, J&K, India. <sup>2</sup>Centre of Research for Development, University of Kashmir 190 006, J&K, India.

#### Received 18 August, 2014; Accepted27 November, 2014

*Crocus sativus* L. is native to Iran and Greece, and is now cultivated largely in Southern Europe, Tibet and other countries. In India, it is mainly cultivated in Kashmir. *C. sativus* is an important medicinal plant with aphrodisiac, antispasmodic, expectorant, anti-diabetic, anti-inflammatory, antioxidant, antidepressant, anticancer and anti-tumor activities. Phytochemical investigations of the species have revealed the presence of a number of important carotenoids especially crocetin and its glycosidic forms such as crocin, picrocrocin and safranal. The genetic origin of *C. sativus* is believed to have occurred by auto-triploidy or by allopolyploidy and *Crocus cartwrightianus* is believed to be its most probable ancestor. World over, saffron shows a declining trend in production and productivity due to high labour cost, lack of variability for major economic traits and poor economic returns. This review focuses on the detailed distribution, chemical composition and the medicinal importance of saffron.

Key words: Crocus sativus, crocin, picrocrocin, safranal, medicinal properties.

#### INTRODUCTION

*Crocus sativus* L. (Family Iridaceae) commonly known as saffron is distributed primarily in the Mediterranean Region and South Western Asia. The safranal (for ordor), picrocrocin (for taste) and crocin (for pigment) components of this geophyte constituting the spice "saffron" are localized in the red stigmatic lobes of the flower (Neghbi et al., 1989; Plessner et al., 1989; Fernandez, 2004). The stigmas (20 - 40 mm) are dark red in color and trumpet shaped, serrated or indented at the distal end and may be isolated or joined in pairs or threes at the end of the style, which is white/yellow in color (Figure 1b, c). It is estimated that, approximately 75,000 crocus blos-

soms or more than 2,00,000 dried stigmas produce just one 1 kg saffron spice and is thus the most expensive spice in the world at around \$500/kg and/or \$40-50/gram (Fernandez, 2007; Melnyk et al., 2010). The stigmas must be handpicked from the delicate blossoms upon opening to preserve the desirable volatile compounds which easily degrade in the presence of light and oxidizing agents (Rau, 1969; Hill, 2004). As a result, the best saffron is usually sold as whole stigmas (not powdered) in air tight containers so as to preserve its integrity. The high value of saffron in the international market makes it the object of frequent adulteration

\*Corresponding author. E-mail: sabiyajaan26@gmail.com / aawani@kashmiruniversity.ac.in.

Author(s) agree that this article remain permanently open access under the terms of the <u>Creative Commons Attribution License</u> <u>4.0 International License</u>



Figure 1. Corm (1a), flower (1b) and stigma (1c) of Crocus sativus L.

and fraud by the growers, traders and other stake holders (Fernandez, 2007; Husaini et al., 2010a). The quality and commercial value of saffron is determined by its coloring power, bitter taste and aroma. These three parameters are certified in the international market following the International Organization for Standardization (ISO 1993). The "Saffron Specifications" and "Saffron Test Methods" issued by the Institute of Standard and Industrial Research Organization of Iran (ISIROI) explain the sampling, packing, labeling and methods of testing of saffron. In India, the Bureau of Indian Standards (BIS) is responsible for setting up guidelines for quality standards of saffron (Husaini et al., 2010a).

Saffron is a geophyte herbaceous plant and is propagated vegetatively by corms (underground. compact, bulb-like, starch-storing organs shrouded in a dense mat of parallel fibres called "corm tunic"). A single corm of 10-15 g weight survives for one season, producing at the end of growing season 6-10 "cormlets" that can grow into new plants in the next season (Figure 1a). The plant is a triploid (X=8; 2n=3X= 24), self and outsterile, mostly male sterile (Mathew, 1977; Ghaffari, 1986; Grilli Caiola, 2005) and is therefore unable to produce seeds. Archeological and historical sources indicate the saffron cultivation as very old dating back to 2500 - 1500 BC, probably originating in Iran, Asia Minor or Greece which later became wide spread in India, China, the Mediterranean Basin and Eastern Europe (Negbi, 1999; Grilli Caiola et al., 2004). The auto-triploid nature and vegetative mode of reproduction of the species renders improvement by conventional breeding very difficult. As the species has spread by vegetative means, it is believed that saffron exists as a single species all over the world.

Recent studies have confirmed that saffron exhibits stable biological traits all over the world and there are no genomic differences (Grilli caiola et al., 2004; Zubor et al., 2004; Fernandez, 2007). The stigmas of saffron have been used from ancient times as a spice in food, as a dye in perfumes and cosmetics preparation and for medicinal purposes (Basker and Negbi, 1983). The present review describes the current status of saffron crop and its medicinal properties so as to have insight interest among young researchers for their possible contributions in promoting this precious crop in the world.

#### Origin and distribution

The name "saffron" is derived from the Arabic word means vellow zafaran which (Winterhalter and Straubinger, 2000), the ancient Greek called it "Koricos" where as Romans used the term "Crocum". In India, this golden spice is known as "Kum Kum" and "Kesar" in Sanskrit and "Koung" in Kashmiri language. It is believed that saffron is being cultivated for about 3,500 years in Egypt and Middle East and during the Middle Age, saffron crop was extended from Middle East to Europe reaching Great Britain in the 14<sup>th</sup> Century (Fernandez, 2004). The detailed archaeological and historical records of occurrence and spread of saffron and its allied species have been reviewed by Grilli Caiola (2010). The centre of origin of C. sativus according to Vavilov (1951) is the Middle East, while other authors suggest Asia Minor or the South-West Greek Islands as its probable area of origin (Tammaro, 1990). According to Negbi (1999), C. sativus was probably selected and domesticated in Crete during the Late Bronze Age.

From here, it spread to India, China and the Middle Eastern countries. From these latter, the Arabs brought saffron to all Mediterranean Europe (Ingram, 1969). Some authors (Alberini, 1990; Winterhalter and Straubinger, 2000) point towards Iran and Kashmir as its origin site from where it has spread to Greek and Roman world. The precise time of introduction of saffron in Kashmir is not known, although evidence from a 12<sup>th</sup> Century book, "Rajatarangini" written by a Kashmir Poet (kalhana), indicates its presence in Kashmir even before

Country	Area ( ha )	Production (Kg)	References
Iran	47,000	160,000	Ehsanzadeh et al., 2004
India	-	8,000-10,000	Fernandez, 2004
Greece	860	4,000-6,000	Fernandez, 2004
Azerbaijan	675	-	Azizbekova and Milyaeva, 1999
Morocco	500	1,000	Ait-Oubahou and El-Otmani, 1999
Spain	200	300-500	Fernandez, 2004
Italy	35	120	NA
France	1	4	Girard and Navarrete, 2005
Turkey	-	10	Thiercelin, 2004
Switzerland	-	0.4	Negbi, 1999

 Table 1. Estimate of saffron world production (Adopted from Gresta et al., 2008).

NA - reference not available.

the reign of King Lalitaditya in 750AD (Husaini et al., 2010b). The genetic origin of C. sativus is believed to have occurred by auto-triploidy from a wild Crocus, probably by fertilization of a diploid unreduced egg cell by a haploid sperm cell or a haploid egg cell by two haploid sperms (Chichiriccò, 1984; Grilli Caiola, 2004, 2005), or by allopolyploidy through the hybridization of Crocus cartwrightianus and Crocus hadriaticus (Castillo et al., 2005). Brighton (1977) in a kariological study and supported by AFLP analysis (Zubor et al., 2004) suggested that possible ancestors of C. sativus are C. cartwrightianus or Crocus thomasii. Evidences from several other workers suggest C. cartwrightianus as the most probable ancestor of C. sativus (Mathew, 1999; Brandizzi and Grilli Caiola, 1998; Grilli Caiola et al., 2004).

Saffron is currently being cultivated more or less intensively in Iran, India, Greece, Spain, Italy, Turkey, France, Switzerland, Israel, Pakistan, Azerbaijan, China, Egypt, Japan, Afghanistan, Iraq and recently in Australia (Table 1). While the world's total annual saffron production is estimated at 205t per year, Iran with more than 47,000 ha, of land under saffron cultivation produces 80% (160t) of this total. Khorasan province in Iran alone accounts for 46,000 ha land and 137t of the total production in Iran (Parviz et al., 2004). The traditional cultivated areas in Europe (Spain, Italy and Greece) are showing a severe declining trend while an enormous increase has been registered in Iran in the last 30 years (Skubris, 1990; Fernandez, 2004). In India saffron is exclusively cultivated in Kashmir division of Jammu and Kashmir State.

Locally known as "Koung" and generally grown on uplands (Karewas), this crop covers about 4% of the total cultivated area of the Kashmir valley and produces 5-6t annually (Husaini et al., 2010a, b). Saffron export from India declined from 9.7t (1998-99) to 8.7 tons (2000-01) associated with a decline in spot price of saffron from Rupees (Rs.) 32,936/kg (\$ 866) in 1997-98 to Rs.17, 500 (\$ 374) in 2004-05 (Nehvi et al., 2007). The total area under this crop and annual production in the state is showing a declining trend over the past more than one decade (Table 2). According to Husaini et al. (2010b), the saffron crop has shown a decrease of 83% in area, 215% in production and 72% in productivity. The major reasons for decline in saffron cultivation and production constraints in the world as well as in J&K state of India are high labour cost, lack of variability for major economic traits, low corm yield, disease susceptibility, low yield of biochemical like safranin, picrocrocin and crocin and above all poor economic returns. The cultivation practices of saffron in Kashmir and the factors responsible for decline in saffron production have been reviewed by Husaini et al. (2010b).

#### Chemical constituents of saffron

Apart from the primary metabolites such as carbohydrates, minerals, fats and vitamins, the Crocus sativus L. contains four major bioactive compounds viz., crocin (Mono-glycosyl polytene esters), crocetin (a natural carotenoid dicarboxylic acid precursor of crocin), picrocrocin (monoterpene glycoside precursor of safranal and product of xeaxanthin degradation) and safranal (Figure 2), all contributing to colour, taste and aroma respectively (Melnyk et al., 2010). According to Sobolev et al. (2014), presence of biologically active compounds such as crocetin, picrocrocin and safranal makes this spice a promising candidate for being a functional food. The hydrophilic carotenoids of saffron which includes crocins constitute about 6-16% of saffron's dry matter depending upon the variety, growing conditions and processing methods (Gregory et al., 2005). The highly water soluble crocins are widely used as a natural food colourant and also act as an antioxidant by guenching free radicals, thus protecting cells and tissues against oxidation (Assimopolou et al., 2005; Soeda et al., 2007; Melnyk et al., 2010). Amongst the other minor components belonging to this class, β-crocetin and y-

Year	Area (ha)	Production (ton)	Yield/productivity (kg ha <sup>-1</sup> )
1996 - 1997	5707	15.96	2.79
1997 - 1998	NA	NA	NA
1998 - 1999	4116	12.88	3.12
1999 - 2000	3997	7.65	1.91
2000 - 2001	2831	3.59	1.26
2001 - 2002	2880	6.52	2.26
2002 - 2003	2742	5.15	1.87
2003 - 2004	3075	4.83	1.57
2004 - 2005	2989	8.85	2.96
2005 - 2006	2928	4.85	1.65
2006 - 2007	2436	9.13	3.74
2007 - 2008	3110	5.06	1.62

Table 2. Area, production and productivity of saffron in Kashmir, India.

Adopted from Husaini et al. (2010) (Sources: Planning Department J&K Government/Directorate of Agriculture Jammu and Kashmir Divisions/Economic Survey 2008 - 09, J&K Government)

NA: Data not available



Crocetin



### Picrocrocin

Safranal

Figure 2. Major chemical constituents of Crocus sativus L. (adopted from Gresta et al., 2008).

crocetin, the mono and dimethyl esters of crocetin respectively and mangi-crocin, an unusual xanthonecarotenoid glycosidic conjugate, have also been identified (Sampathu et al., 1984; Ghosal et al., 1989; Fernandez, 2004). The picrocrocin which is the second most abundant component (1-13% of saffrons dry matter) is a colourless glycoside and is considered the main bitter principle of saffron, even though other components, such as flavonoids are also responsible for saffron's bitterness (Alonso et al., 2001; Carmona and Alonoza, 2004). Picrocrocin like other members of the crocin family is derived from the enzymatic degredation of zeaxanthin; in turn, the natural de-glycosylation of picrocrocin gives safranal (Sampathu et al., 1984; Pfander and Schurteberger, 1982) which is the main volatile component of saffron, responsible for the particular aroma of this spice. The safranal represents approximately 30-37% of essential oil and 0.001 to 0.006% of dry matter (Carmona et al., 2007; Maggi et al., 2009). Besides its aromatic potential, safranal has antioxidant potential (Kanakis et al., 2007) and cytotoxic effect on certain cancer cells (Escribano et al., 1996). There are also other typical volatile components of saffron, all possessing the same skeleton of safranal and like this, are considered to derive from picrocrocin (Melnyk et al., 2010), even though the recent discovery of several new glycosides suggests that picrocrocin is not the soluble glycosidic aroma precursor in saffron (Straubinger et al., 1998; Carmona et al., 2006). The extraction and purification protocol for various chemical and volatile constituents of saffron are available in the literature (Tarantilis et al., 1994; Tarantilis and Polissiou, 1997; Lozano et al., 2000, Zareena et al., 2001).

Several minor components have also been isolated from stigmas and other plant parts, mainly petals and corms. Terpenoids such as crocusatins present in stigmas and petals and showing a significant antityrosinase activity, are among the most recovered components (Li and Wu, 2002, 2004). To the same class of substances namely terpenoids, belong several glycosidic derivatives which are considered as the precursors of volatile saffron components alternative to picrocrocin (Straubinger et al., 1997, 1998). Moreover, a series of flavonoids, all glycosidic derivatives of kaempferol, have recently been characterized in the stigmas of saffron; these polyphenols probably concur together with picrocrocin to produce the bitter taste of saffron (Carmona et al., 2007). Other secondary metabolites from C. sativus include anthraguinones and an anthocyanin (Saito et al., 1960; Gao et al., 1999), isolated from corms and petals, respectively.

The chemical composition and concentration of various metabolites in saffron vary from one geographical region to other. Several analytical techniques are available to differentiate saffron samples of different origin based on their chemical composition (Zalacain et al., 2005; Zougagh et al., 2006; Maggi et al., 2009, 2011; Yilmaz et al., 2010). Recently, Sobolev et al. (2014) proposed a microwave assisted NMR based analytical protocol for recovery of metabolites showing significant differences among geographically different saffron extracts.

#### Medicinal importance of saffron

Saffron (*C. sativus* L.) has been cultivated from time immemorial for its stigmas, which not only comprise a highly valued spice but also have various therapeutic uses (Sampathu et al., 1984). It is used mainly as a dye in industry, as a spice in cooking, as a food colorant and as a component of drugs and perfumes (Mathew, 1982; Basker and Negbi, 1983; Behnia et al., 1999). Saffron has been used as a drug to treat various human health conditions such as coughs, stomach disorders, colic, insomnia, chronic uterine hemorrhage, scarlet fever, smallpox, colds, asthma and cardiovascular disorders (Giaccio, 1990; Winterhalter and Straubinger, 2000; Abdullave, 2003). It has been shown that saffron is a protective agent against chromosomal damage (Prem kumar et al., 2001). Saffron can also be used to help clear up sores and to reduce the discomfort of teething infants (Abdullaev and Espinosa- Agurre, 2004).

Among the secondary metabolites present in saffron, the ester derivatives of crocetin, together with safranal, are nowadays the most studied compounds to evaluate their biological activity. Recent data shows that saffron possesses tyrosinase inhibitory (Li and Wu, 2002, 2004), anticonvulsant (Hosseinzadeh and Younesi Hani, 2002), mutagenic (Abdullave and Espinosa-Aguirre, 2004), cytotoxic and antigenotoxic effects (Abdullaev et al., 2003). It has also anti-amyloidogenic activity against Alzheimer's disease (Papandreou et al., 2006); antiinflammatory (Hosseinzadeh and Younesi, 2002) and blood pressure reducing (Fatehi et al., 2003) effects. Crocin extracts from saffron have been used for the treatment of nervous, cardiovascular and respiratory systems (Abe and Saito, 2000; Abdullaev, 2002). Components of saffron extract have been found to play a role in management of mental disorders and also act as antidepressant agents (Lechtenberg et al., 2008; Basti et al., 2007). It has been found that the treatment with saffron extracts is not associated with sexual dysfunction in humans, a side effect often encountered with antidepressant drugs (Modabbernia et al., 2012; Kashani et al., 2013).

Recently, saffron extract has been successfully tested as an anticancer agent (Abdullaev, 2007) as well as against mental disorders. Cancer chemopreventive and tumoricidal properties of saffron extracts have been reported by several workers following in vitro and in vivo assays with encouraging results (Abdullaev, 2002; Ochiai et al., 2004; Ahmad et al., 2005; Hosseinzadeh et al., 2005; Magesh et al., 2006). According to Hartwell (1982) saffron extracts have been used against different kinds of tumors and cancers (liver, spleen, kidney, stomach and uterus tumors) in ancient times. Anti-tumor effect of saffron on different malignant cells in some model animals has also been reported (Abdullaev, 2004). Very recently, De Monte et al. (2014) studied the inhibitory activities of two natural components of C. sativus viz. crocin and safranal as well as some newly designed components derived from chemical modifications of safranal on the human monoamine oxidases (hMAO-A and hMAO-B- the two important enzymes which are targets for the treatment of neuropsychiatric and neurodegenerative diseases). Their results confirmed crocin as a relatively weak inhibitor of hMAO, while as safranal was not found as hMAO inhibitor indicating that hMAO are probably not targets of crocin and safranal. The designed chemical derivatives of safranal, however, displayed much improved inhibitory activities against both hMAO enzymes. The synthetic derivatives could thus

 Table 3. Major biological functions attributed to saffron and its chemical constituents.

Activity	Saffron constituents tested	Reference
Prevention of gastric disorder	Saffron crude extract	Inoue et al. (2005)
5	Ethanolic saffron extract	Kianbakht and Mozaffari (2009).
Prevention of stomach ulcer	Crocin	Xu et al. (2009)
Digestion enhancement	Aqueous saffron extract	Nabavizadeh et al. (2009)
Anticancer function and cytotoxic effects on tumor cells	Ethanolic saffron extract Crocin, crocetin, safranal and picrocrocin	Tavakkol-Afshari et al (2008) Escribano et al. (1996) Garcia-Olmo et al. (1999) Abdullaev (2002) Mousavi et al. (2009)
Tumor inhibition	Crocin Saffron Crocetin	Garcia-Olmo et al. (1999) Salomi et al. (1991) Wang et al. (1996)
Cardiovascular health promotion Anti-atherosclerosis	Crocin Crocetin	He et al. (2005) He et al. (2007) Sheng et al. (2006)
Prevention of insulin resistance	Crocetin	Xi et al. (2007)
Anti-depression activities	Capsulated ethanolic saffron extract Saffron petal extract Aqueous and ethanolic saffron extract	Akhondzadeh et al. (2005) Akhondzadeh et al. (2007) Moshiri et al. (2006) Hosseinzadeh et al. (2004)
Premenstrual syndrome (PMS) treatment	Capsulated ethanolic saffron extract	Agha-Hosseini et al. (2008)
Detrimental health effects Nausea, vomiting, uterus bleeding, abortion	Saffron	Schimidt et al. (2007) Lucas et al. (2001)

Modified and adopted from Melnyk et al. (2010).

prove novel hMAO inhibitors for clinical management of psychiatric and neurodegenerative disorders. A detailed review by Melnyk et al. (2010) highlights major biological functions attributed to different constituents of saffron (Table 3).

#### PROSPECTS OF GENETIC IMPROVEMENT

Saffron's high price is due to the much direct labour required for its cultivation, harvesting and handling (Fernandez, 2004). In recent past, saffron cultivation and production has shown a declining trend due to high labour cost, low economic returns and very short and laborious flower picking period. There is an urgent need for increasing saffron production and quality to cope with an increasing demand of this spice in the market. This can be achieved by putting in more efforts on genetic improvement of the crop with main focus on producing more flowers per plant, flowers with a higher number of stigmas, increasing stigmas size or stigmas with an increased amount of dye and aroma (Fernandez, 2004). Due to triploid behavior, the chances of crop improvement by conventional methods like hybridization are not possible (Basker and Negbi, 1983). The utilization of spontaneous variability in the natural population which

is due to genetic and environmental factors and other non-conventional approaches of crop improvement offer tremendous scope for saffron improvement (Estilai, 1978; Dhar et al., 1988; Nehvi, 2003). Several workers in recent years have attempted mutation breeding technique for induction of genetic variability followed by selection and multiplication of mutant clones (Nehvi et al., 2010). The use of mutagenesis could enhance the genetic base of the crop species so as to offer chances of selection for elite genotypes particularly with respect to stigmatic and corm characteristics. The preliminary results of induced genetic variability through gamma irradiation and induction of polyploidy through colchinization are, however, not satisfactory and probably would require further work (Akhund-Zade and Mazaferova, 1975; Khan, 2004; Zaffar et al., 2004). Creation of saffron germplasm banks, improvement in cultivation techniques, supply of quality plant material and development of quality evaluation methods are some important measures to be considered while dealing with enhancing saffron productivity and its usage.

#### CONCLUSION

Saffron cultivation has been neglected for many decades

by farmers, who have relegated it to adverse soil and climate conditions, and by research, which has led to a lack of innovation. The chemical profile of saffron and its medicinal and cultural properties makes it a golden spice and there is an urgent need of attention on scientific community to focus their research on genetic improvement of this precious crop. Increase in saffron production and quality can be achieved by means of plants with more flowers per plant, flowers with a higher number of stigmas, increased stigma size or stigmas with a greater amount of dye and aroma. The sterility of saffron limits the application of conventional breeding approaches for its further improvement. Induced variability by physical and chemical mutagens can be considered a viable option for improvement in saffron yield, even if no significant results have been achieved as yet.

#### **Conflict of Interests**

The author(s) have declared that there is no conflict of interests.

#### ACKNOWLEDGEMENT

The first author is grateful to Department of Biotechnology, Government of India for providing financial assistance as JRF in the project entitled "Induction of Variability for Genetic Improvement of Kashmir Saffron".

#### REFERENCES

- Abdullaev F (2003). Crocus sativus against cancer. Arch. Med. Res. 34:354-354.
- Abdullaev F (2007). Biological properties and medicinal use of saffron (*Crocus sativus*). Acta Hort. 739:339-345.
- Abdullaev FI (2002). Cancer chemopreventive and tumoricidal properties of saffron (*Crocus sativus* L.). Exp. Biol. Med. 227:20-25.
- Abdullaev FI (2004). Antitumor effect of saffron (*Crocus sativus* L.). Overview and prespectives. Acta Hort. 650:491-499.
- Abdullaev FI, Espinosa-Aguirre JJ (2004). Biomedical properties of saffron and its potential use in cancer therapy and chemoprevention trials. Cancer Detect. Prev. 28:426-432.
- Abdullaev FI, Riveron-Negrete L, Caballero-Ortega H, Herdandez JM, Perez-Lopez I, Pereda-Miranda R, Espinosa-Aguirre JJ (2003). Use of *in vitro* assays to assess the potential antigenotoxic and cytotoxic effects of saffron (*Crocus sativus* L.). Toxicol. Vitro 17:731-736.
- Abe K, Saito H (2000). Effects of saffron extract and its constituent crocin on learning behavior and long- term potentiation. Phytother. Res. 14:149-152.
- Agha-Hosseini M, Kashani L, Aleyaseen A, Ghoreishi A, Rahmanpour H, Zarrinara AR (2008). *Crocus sativus* L. (saffron) in the treatment of premenstrual syndrome: A double-blind, randomized and placebocontrolled trial. BJOG — An International J. Obstet. Gynaecol. 115(4):515-519.
- Ahmad AS, Ansari MA, Ahmad M, Muzamil SS, Yousuf S, Hoda MN, Islam F (2005). Neuroprotection by crocetin in a hemi-parkinsonian rat model. Pharmacol. Biochem. Behav. 81:805-813.
- Akhondzadeh S, Tahmacebi-Pour N, Noorbala A, Amini H, Fallah-Pour H, Jamshidi A (2005). *Crocus sativus* L. in the treatment of mild to moderate depression: A double blind, randomized and placebo controlled trial. Phytother. Res. 19:148-151.

- Akhondzadeh Basti A, Moshiri E, Noorbala A, Jamshidi A, Abbasi SH, Akhondzadeh S (2007). Comparison of petal of Crocus sativus L. and fluoxetine in the treatment of depressed outpatients: A pilot doubleblind randomized trial. Prog. Neuro-Psychopharmacol. Biol. Psychiatry 31(2):439-442.
- Akhund-Zade IM, Muzaferova RS (1975). Study of the effectiveness of gamma irradiations on the saffron. Radiobiol. 15:319-322.
- Alberini M (1990). Saffron e colore. Lo Zafferano. Proceedings of International Conference on Saffron (*Crocus sativus* L.). L' Aquila, Italy, pp. 39-46.
- Alonso GL, Salinas MR, Garijo J, Sanchez-Fernadez MA (2001). Composition of crocins and picrocrocin from Spanish saffron (*Crocus sativus* L.). J. Food Qual. 24(3):219-233.
- Basker D, Negbi M (1983). The use of Saffron. Eco. Bot. 37:228-236.
- Basti AA, Moshiri E, Noorbala AA, Jamshidi AH, Abbasi SH, Akhondzadeh S (2007). Prog. Neuro-psychopharmacol. Biol. Psychiatry 31:439-442.
- Behnia MR, Estilai A, Ehdaie B (1999). Application of fertilizer for increased saffron yield. J. Agric. Crop Sci. 182:9-15.
- Bouvier F, Suire C, Mutterer J, Camara B (2003) not cited. Oxidative remodeling of chromoplast carotenoids: identification of the carotenoid dioxygenase CsCCD and CsZCD genes involved in Crocus secondary metabolite biogenesis. Plant Cell. 15:47-62.
- Brandizzi F, Grilli Caiola M (1998). Flow cytometric analysis of nuclear DNA in *Crocus sativus* and allies (Iridacee). Plant. Syst. Evol. 211:149-154.
- Carmona M, Alonzo GL (2004). A new look at saffron: mistakes beliefs. Proceedings of the First International Symposium on Saffron Biology and Biotechnology. Acta Hort. 650:373-391.
- Carmona M, Sánchez AM, Ferreres F, Zalacain A, Tomás-Berberán F, Alonso GL (2007). Identification of the flavonoid fraction in saffron spice by LC/DAD/MS/MS: comparative study of samples from different geographical origin. Food Chem. 100:445-450.
- Carmona M, Zalacain A, Sanchez AM, Novella JL, Alonso GL (2006). Crocetin esters, picrocrocin and its related compounds present in *Crocus sativus* stigmas and *Gardenia jasminoides* fruits. Tentative identification of seven new compounds by LC-ESI-MS. J. Agric. Food Chem. 54:973-979.
- Carmona M, Zalacain A, Salinas MR, Alonso GL (2007). A new approach to saffron aroma. Crit. Rev.Food Sci. Nutr. 47:145-159.
- Castillo R, Fernandez JA, Gomez-Gomez L (2005). Implications of carotenoid biosynthetic genes in apocarotenoid formation during the stigma development of *Crocus sativus* and its closer relatives. Plant Physiol. 139:674-689.
- Chichiriccò G (1984). Karyotype and meiotic behaviour of the triploid *Crocus sativus* L. Caryologia 37:233-239.
- De Monte C, Carradori S, Chimenti P, Secci D (2014). New insights into the biological properties of Crocus sativus L. chemical modifications, human monoamine oxidases inhibition and molecular modeling studies. Eur. J. Med. Chem. 82:164-171.
- Dhar AH, Sapru R, Rebha K (1988). Studies on saffron in Kashmir. Variation in natural population on cytological behaviour. Crop Improv. 15(1):48-52.
- Escribano J, Alonso GL, Coca-Prados M, Fernandez JA (1996). Crocin, safranal and picrocrocin from saffron (*Crocus sativus* L.) inhibit the growth of human cancer cells in vitro. Cancer Lett. 100:23-30.
- Estilai A (1978). Variability in saffron (*Crocus sativus* L.). Experientia 34:725-727.
- Fatehi M, Rashidabady T, Fatehi- Hassanabad Z (2003). Effects of *C. sativus* petals extract on rat blood pressure and on responses induced by electrical field stimulation in the rat isolated vas deferens and guinea pig ileum. J. Ethnopharmacol. 84:199-203.
- Fernandez JA (2004). Biology, biotechnology and biomedicine of Saffron.Recent Res. Dev. Plant Sci. 2:127-159.
- Fernandez JA (2007). Genetic resources of saffron and allies (*Crocus* spp.). Acta Horticulturae. 739:167-185.
- Gao WY, Li YM, Zhu DY (1999). New anthraquinones from the sprout of *Crocus sativus*, Acta Bot. Sin. 41:531-533.
- Garcia-Olmo DC, Riese HH, Escribano J, Ontanon J, Fernandez JA, Atienzar, M (1999). Effects of long-term treatment of colon adenocarcinoma with crocin, a carotenoid from saffron (*Crocus sativus* L.): An experimental study in the rat. Nutr. Cancer 35(2):120-126.

- Ghaffari SM (1986). Cytogenetic studies of cultivated *Crocus sativus* (Iridacee). Plant Syst. Evol. 153:199-204.
- Ghosal S, Singh SK, Battacharya SK (1989). Mangicrocin, and adaptogenicxanthone-carotenoid glycosidic conjugate from saffron. J. Chem. Res. 3:70-71.
- Giaccio M (1990). Components and features of saffron. Proceedings of the international conference on Saffron (ICS'90), L'fAquila, Italy. 135-148.
- Gregory MJ, Menary RC, Davies NW (2005). Effect of drying temperature and air flow on the production and retention of secondary metabolites in saffron. J. Agric. Food Chem. 53(15): 5969-5975.
- Grilli Caiola M (2004). Saffron reproductive biology. Acta Hort. 650:25-37.
- Grilli Caiola M (2005). Embryo origin and development in *Crocus* sativus L. (Iridacee). Plant Biosyst. 139:335-343.
- Grilli CM, Antonella C (2010). Looking for saffron's (*Crocus sativus* L.) parents. . Funct. Plant Sci. Biotechnol. 4(2):1-14, Global Science Books.
- Grrili CM, Caputo P, Zanier R (2004). RAPD analysis in *Crocus sativus* L. accessions and related Crocus species. Biol. Plant. 48:375-380.
- Hartwell JL (1982). Plants used against cancer: A survey. Quaterman Publications, Lawrence, CA.
- He SY, Qian ZY, Tang FT, Wen N, Xu GL, Sheng L (2005). Effect of crocin on experimental atherosclerosis in quails and its mechanisms. Life Sci. 77(8):907-921.
- He SY, Qian Z. Y, Wen N, Tang FT, Xu GL, Zhou CH (2007). Influence of crocetin on experimental atherosclerosis in hyperlipidemic-diet quails. Eur. J. Pharmacol. 554(23):191-195.
- Hill T (2004). The Contemporary Encyclopedia of Herbs and Spices: Seasonings for the Global Kitchen (1st ed.), Wiley, <u>ISBN 978-0-471-21423-6</u>.
- Hosseinzadeh H, Sadeghnia HR, Ziaee TD (2005). Protective effect of aqueous saffron extract (*Crocus sativus* L.) and crocin, its active constituent, on renal ischemia-reperfusion-induced oxidative damage in rats, J. Pharm. Sci. 8:387-393.
- Hosseinzadeh H, Yonesi HM (2002). Antinociceptive and antiinfilamatory effects of *Crocus sativus* L. stigma and petal extracts. BMC. Pharmacol. 2:7-7.
- Hosseinzadeh H, Karimi G, Niapoor M (2004). Antidepressant effect of *Crocus sativus* L. stigma extracts and their constituents, crocin and safranal, in mice. Proceedings of the 1st International Symposium on Saffron Biology and Biotechnology, 650:435-445.
- Husaini AM, Kamili AN, Wani MH, da Silva JAT, Bhat GN (2010a). Sustainable saffron (*Crocus sativus* Kashmirianus) production: Technological and Policy interventions for Kashmir. Functional Plant Science and Biotechnology, Global Science Books. 4(2):116-127
- Husaini AM, Hassan B, Ghani MY, da Silva JAT, Kirmani NA (2010b). Saffron (*Crocus sativus* Kashmirianus) cultivation in Kashmir: Practices and problems. Funct. Plant Sci. Biotechnol. 4(2): 108-115, Global Science Books.
- Inoue E, Shimizu Y, Shoji M, Tsuchida H, Sano Y, Ito C (2005). Pharmacological properties of N-095, a drug containing red ginseng, polygala root, saffron, antelope horn and aloe wood. Am. J. Chin. Med. 33(1):49-60.
- Ingram JS (1969). Saffron (Crocus sativus L.). Trop. Sci. 11:177-184.
- International Organization for Standardization (ISO) (1993). Saffron (Crocus sativus Linnaeus). Pt 1: Specifications. Pt 2: Test Methods; ISO: Geneva, Switzerland.
- Kanakis CD, Tarantilis PA, Tajmir-Riahi HA, Polissiou MG (2007). Crocetin, dimethylcrocetin, and safranal bind human serum albumin: Stability and antoxidative properties. J. Agric. Food Chem. 55(3):970-977.
- Kashani L, Raisi F, Saroukhani S, Sohrabi H, Modabbernia A, Nasehi AA, Jamshidi A, Ashrafi M, Mansouri P, Ghaeli P, Akhondzadeh S (2013). Hum. Psychopharmacol. Clin. Exp. 28:54-60.
- Kianbakht S, Mozaffari K (2009). Effects of saffron and its active constituents, crocin and safranal on prevention of indomethacin induced gastric ulcers in diabetic and non diabetic rats. J. Med. Plants 8(5):30-38.
- Khan IA (2004). Induced mutagenic variability in saffron (*Crocus sativus* L.), Acta Hort. 650:281-283.

- Lechtenberg M, Schepmann D, Niehues M, Hellenbrand N, Wünsch B, Hensel A (2008). Planta Med. 74:764-772.
- Li CY, Wu TS (2002). Constituents of the Stigmas of *Crocus sativus* and Their Tyrosinase Inhibitory Activity. J. Nat. Prod. 65:1452-1456.
- Li CY, Wu TS (2004). Antityrosinase principles and constituents of the petals of *Crocus sativus*, J. Nat. Prod. 6:437-440.
- Lozano P, Delgado D, Gomez D, Rubio M, Iborra JL (2000). A nondestructive method to determine the safranal content of saffron (Crocus sativus L.) by supercritical carbon dioxide extraction combined with high-performance liquid chromatography and gas chromatography. J. Biochem. Biophys. Methods 43(13):367-378.
- Lucas CD, Hallagan JB, Taylor SL (2001). The role of natural color additives in food allergy. Adv. Food Nutr. Res. 43:195-216.
- Magesh V, Singh JPV, Selvendiran K, Ekambaram G, Sakthisekaran D (2006). Antitumour activity of crocetin in accordance to tumor incidence, antioxidant status, drug metabolizing enzymes and histopathological studies. Mol. Cell. Biochem. 287:127-135.
- Mathew B (1977). *Crocus sativus* L. and its allies (Iridaceae). Plant Syst. Evol. 128:89-103.
- Mathew B (1982). The Crocus. A revision of the genus Crocus (Iridaceae). Batsford, B.T.Ltd., London.
- Mathew B (1999). Botany, taxonomy and cytology of *Crocus sativus* L. and its allies. In: Negbi M (Ed.), Saffron, Harwood Academic Publishers, Amsterdam.
- Maggi L, Carmona M, del Campo CP, Kanakis CD, Anastasaki E, Tarantilis PA (2009). Worldwide market screening of saffron volatile composition. J. Sci. Food Agric. 89(11):1950-1954.
- Maggi L, Carmona M, Kelly SD, Marigheto N, Alonso GL (2011). Geographical origin, differentiation of saffron spice (*Crocus sativus* L. stigmas)—Preliminary investigation using chemical and multi-element (H, C, N) stable isotope analysis. Food Chem. 128:543-548.
- Moshiri E, Akhondzadeh Basti A, Noorbala AA, Jamshidi AH, Abbasi SH, Akhondzadeh S (2006). Crocus sativus L. (petal) in the treatment of mild-to moderate depression: A double-blind, randomized and placebo-controlled trial. Phytomed. 13(9-10):607-611.
- Modabbernia A, Sohrabi H, Nasehi AA, Raisi F, Saroukhani S, Jamshidi A, Tabrizi M, Ashrafi M, Akhondzadeh S (2012). Effect of saffron on fluoxetine-induced sexual impairment in men: randomized doubleblind placebo-controlled trial. Psychopharmacol. 223:381-388.
- Melnyk JP, Wang S, Marcone MF (2010). Chemical and biological properties of the world's most expensive spice: Saffron. Food Res. Int. 43:1981-1989.
- Mousavi SH, Tavakkol-Afshari J, Brook A, Jafari-Anarkooli I (2009). Role of caspases and Bax protein in saffron-induced apoptosis in MCF-7 cells. Food Chem. Toxicol. 47(8):1909-1913.
- Nabavizadeh F, Salimi E, Sadroleslami Z, Karimian SM, Vahedian J (2009). Saffron (*Crocus sativus*) increases gastric acid and pepsin secretions in rats: Role of nitric oxide (NO). Afr. J. Pharm. Pharmacol. 3(5):181-184.
- Negbi M (1999). Saffron cultivation: past, present and future prospects. In: Negbi M (ed.): Saffron, *Crocus sativus* L. Harwood Academy Publ. Australia, p.1-18.
- Nehvi FA (2003). Problems and prospects of saffron improvement in India. Proc. International Seminar on Industrial use of biotechnology. 27th September to 1<sup>st</sup> October, 2003. Islamic Republic of Iran.
- Nehvi FA, Khan MA, Lone AA, Makhdoomi MI, Wani SA, Yousuf V (2010). Effect of radiation and chemical mutagens on variability in saffron (*Crocus sativus* L.). Acta Hort. 850:67-74.
- Nehvi FA, Wani SA, Dar SA, Makhdoomi MI, Allie BA, Mir ZA (2007). Biological interventions for enhancing saffron productivity in kashmir. Acta Horticulturae. 739:25-31.
- Ochiai T, Ohno S, Soeda S, Tanaka H, Shoyama Y, Shimeno H (2004). Crocin prevents the death of rat pheochromyctoma (PC- 12) cells by its antioxidant effects stronger than those of α-tocopherol. Neurosci. Lett. 362:61-64.
- Papandreou MA, Kanakis CD, Polissiou MG, Efthimiopoulos S, Cordopatis P, Margarity M, Lamari FN (2006). Inhibitory activity on amyloid-β-aggregation and antioxidant properties of *Crocus sativus* stigmas extract and its crocin constituents. J. Agric. Food Chem. 54:8162-8168.
- Parviz E, Abbas A, Yadollahi A, Maibodi MM (2004). Productivity, growth and quality attributes of 10 Iranian saffron accessions under

climatic conditions of Chahar-Mahal Bakhtiari central Iran. Acta Hort. 650:183-88.

- Pfander H, Schurteberger H (1982). Biosynthesis of C20-carotenoids in *Crocus sativus*, Phytochem. 21:1039-1042.
- Plessner O, Negbi M, Ziv M, Basker D (1989). Effect of temperature on the flowering of the saffron crocus (*Crocus sativus* L.): induction of hysteranthy. Israel J. Bot. 38:1-7.
- Prem kumar K, Abraham SK, Santhiya ST, Gopinath PM, Ramesh A (2001). Inhibition of genotoxicity by saffron (Crocus sativus L.) in mice. Drug Chem. Toxicol. 24:421-428.
- Rau SR (1969). The Cooking of India, Foods of the World. Time-Life Books, ISBN 978-0-8094-0069-0.
- Saito N, Mitsui S, Hayashi K (1960). Anthocyanins. XXXIII. Delphin, the anthocyanin of medical saffron and its identity with hyacin by paper chromatography of partial hydrolyzates. Proc. Jpn. Acad. 36:340-345.
- Salomi MJ, Nair SC, Panikkar KR (1991). Inhibitory effects of Nigella sativa and saffron (Crocus sativus) on chemical carcinogenesis in mice. Nutr. Cancer 16(1):67-72.
- Sampathu SR, Shivashankar S, Lewis YS (1984). Saffron (*Crocus sativus* L.): Cultivation, processing, chemistry and standardization. Crit. Rev. Food Sci. Nutr. 20:123-157.
- Schmidt M, Betti G, Hensel A (2007). Saffron in phytotherapy: Pharmacology and clinical uses. Wiener Medizinische Wochenschrift 157(13-14):315-319.
- Sheng L, Qian Z, Zheng S, Xi L (2006). Mechanism of hypolipidemic effect of crocin in rats: Crocin inhibits pancreatic lipase. Eur. J. Pharmacol. 543(1-3):116-122.
- Soeda S, Ochiai T, Shimeno H, Saito H, Abe K, Tanaka H (2007). Pharmacological activities of crocin in saffron. J. Nat. Med. 61(2):102-111.
- Straubinger M, Bau B, Eckstein S, Fink M, Winterhalter P (1998). Identification of novel glycosidic aroma precursors in saffron (*Crocus* sativus L.). J. Agric. Food Chem. 46:328-3243.
- Straubinger M, Jezussek M, Waibel R, Winterhalter P (1997). Novel glycosidic constituents from saffron, J. Agric. Food Chem. 45:1678-1681.
- Tammaro F (1990). Crocus sativus L. cv. Piano di Navelli (L'Aquila saffron): environment, cultivation, morphometric characteristics, activeprinciples, uses. In: Tammaro F, Marra L (Eds.), Proceedingsof the international conference on saffron (Crocus sativusL.), L'Aquila, pp. 47-57.
- Tarantilis PA, Polissiou MG (1997). Isolation and identification of the aroma components from saffron (*Crocus sativus*). J. Agric. Food Chem. 45(2):456-462.

- Tarantilis PA, Polissiou M, Manfait M (1994). Separation of picrocrocin, cis-transcrocins and safranal of saffron using high-performance liquid chromatography with photodiode-array detection. J. Chromatogr. A 664(1):55-61.
- Tavakkol-Afshari J, Brook A, Mousavi SH (2008). Study of cytotoxic and apoptogenic properties of saffron extract in human cancer cell lines. Food Chem. Toxicol. 46(11):3443-3447.
- Vavilov NI (1951). The origin, variation, immunity and breeding of cultivated plants, The Cronica Botanica, Co., Waltham, Mass.
- Wang CJ, Cheng TC, Liu JY, Chou FP, Kuo ML, Lin JK (1996). Inhibition of protein kinase C and proto-oncogene expression by crocetin in NIH/3T3 cells. Mol. Carcinogenesis 17(4):235-240.
- Winterhalter P, Straubinger M (2000). Saffron: Renewed interest in an ancient spice, Food Rev. Int. 16:39-59.
- Xi L, Qian Z, Xu G, Zheng, S, Sun S, Wen N (2007). Beneficial impact of crocetin, a carotenoid from saffron, on insulin sensitivity in fructose-fed rats. Journal of Nutritional Biochemistry 18(1):64-72.
- Xu GL, Li G, Ma HP, Zhong H, Liu F, Ao GZ (2009). Preventive effect of crocin in inflamed animals and in LPS-challenged RAW 264.7 cells. J. Agric. Food Chem. 57(18):8325-8330.
- Yilmaz A, Nyberg NT, Molgaard P, Asili J, Jaroszewski JW (2010). <sup>1</sup>H NMR metabolic fingerprinting of saffron extracts. Metabolomics 6:511-517.
- Zaffar G, Wani SA, Anjum T, Zeerak NA (2004). Colchicine induced variability in saffron. Acta Hort. 650:277-280.
- Zalacain Á, Ordoudi SA, Díaz-Plaza EM, Carmona M, Blázquez I, Tsimidou MZ, Alonzo GL (2005). Near-Infrared spectroscopy in saffron quality control: Determination of chemical composition and geographical origin. J. Agric. Food Chem. (53):9337-9341.
- Zareena AV, Variyar PS, Gholap AS, Bongirwar DR (2001). Chemical investigation of gamma-irradiated saffron (*Crocus sativus* L.). J. Agric. Food Chem. 49(2):687-691.
- Zougagh M, Ríos A, Valcárcel M (2006). Determination of total safranal by *in situ* acid hydrolysis in supercritical fluid media: Application to the quality control of commercial saffron. Anal. Chim. Acta. 578:17-121.
- Zubor AA, Suranyi G, Gyori Z, Borbely G, Prokisch J (2004). Molecular biological approach of the systematic of *Crocus sativus* L. and its allies. Acta Hort. 650:85-93.